Novel SCC mutation in a patient of Mexican descent with sex reversal, salt-losing crisis and adrenal failure

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Abstract

Congenital adrenal hyperplasia (CAH) is caused by mutations in cytochrome P450 side chain cleavage enzyme (CYP11A1 and old name, SCC). Errors in cholesterol side chain cleavage by the mitochondrial resident CYP11A1 results in an inadequate amount of pregnenolone production. This study was performed to evaluate the cause of salt-losing crisis and possible adrenal failure in a pediatric patient whose mother had a history of two previous stillbirths and loss of another baby within a week of birth. CAH can appear in any population in any region of the world. The study was conducted at Memorial University Medical Center and Mercer University School of Medicine. The patient was admitted to Pediatric Endocrinology Clinic due to salt-losing crisis and possible adrenal failure. The patient had CAH, an autosomal recessive disease, due to a novel mutation in exon 5 of the CYP11A1 gene, which generated a truncated protein of 286 amino acids compared with wild-type protein that has 521 amino acids (W286X). Although unrelated, both parents are carriers. Mitochondrial protein import analysis of the mutant CYP11A1 in steroidogenic MA-10 cells showed that the protein is imported in a similar fashion as observed for the wild-type protein and was cleaved to a shorter fragment. However, mutant’s activity was 10% of that obtained for the wild-type protein in non-steroidogenic COS-1 cells. In a patient of Mexican descent, a homozygous CYP11A1 mutation caused CAH, suggesting that this disease is not geographically restricted even in a homogeneous population.

Learning points:
• Novel mutation in CYP11A1 causes CAH;
• This is a pure population from Central Mexico;
• Novel mutation created early truncated protein.

Background

Protein targeting to a specific cellular compartment depends on two principal criteria: (1) the signal sequence and (2) maintenance of protein folding. Incorrect folding of metabolic enzymes present in the steroidogenic pathway leads to the reduced production of steroids necessary for mammalian development (1). There are many genes involved in the steroidogenic pathway, and a mutation in one has the potential to negatively impact the production...
of cortisol and induce adrenal enlargement, a condition known as congenital adrenal hyperplasia (CAH). A common feature in these mutations is the development of salt-losing crisis and ambiguous genitalia. Although it may be challenging to identify the gene responsible for reduced or low baseline steroid synthesis and subsequently ambiguous genitalia (2), the problem often starts with the availability of first steroid synthesized, pregnenolone, because the synthesis of all steroids depends on (i) cholesterol flow into the mitochondria and (ii) cleavage of the side chain of cholesterol to generate pregnenolone.

The steroidogenic acute regulatory protein (StAR) fosters cholesterol from the outer to inner mitochondrial membrane; thus, StAR mutations that impact this activity result in low levels of pregnenolone (2, 3). Although some mutations are more common in some specific ethnicities, StAR mutations are present in any ethnicity independent of the country of origin. In addition, mutations in the cytochrome P450 side chain cleavage enzyme (CYP11A1) gene may render the protein inactive and result in low pregnenolone synthesis (3, 4). Thus, a full-term pregnancy is likely not to have any disorder in progesterone synthesis due to mutant CYP11A1. In contrast, low levels of progesterone and miscarriage may be due to CYP11A1 mutations that impact pregnenolone to progesterone conversion.

In addition to the correct folding, the activity of CYP11A1 depends on ferrodoxin, ferrodoxin reductase and NADPH, which are associated with several mitochondrial-resident proteins, generating a large molecular weight complex (5, 6). Here, we report for the first time a de novo mutation in CYP11A1 that causes CAH identified in an infant of Mexican descent. The point mutation in exon 5 resulted in a truncated protein that is processed into the mitochondria in a fashion similar to that observed for wild-type CYP11A1. Both parents are carriers of this mutation, which is likely responsible for two previous abortions and the death of a previous newborn. Thus, although CAH is considered autosomal, it may also appear independent of genetic origin and as a result of the founder effect. We overexpressed this mutant CYP11A1 and determined its activity, folding and expression and compared it with the wild-type protein.

**Case presentation**

The proband was born by emergency C-section due to an approximately 10% placental abruption at 36 weeks and 1 day gestational age with a birth weight of 2416 g (26–50th percentile-for-gestational age), birth length of 46.8 cm (26–50th percentile-for-gestational age) and a head circumference of 34.5 cm (76–90th percentile-for-age). APGARS were 5 at 1 min and 7 at 5 min and 9 at 10 min. Prenatal ultrasound revealed a nuchal fold. Amniocentesis was refused by the mother.

The parents were originally from Central Mexico. A pedigree analysis is presented in Fig. 1A. At the time of proband’s birth, the mother was G4, P4 A2. The prior live-born infant, reportedly appeared female, was born at 23 weeks and 5 days gestational age with a weight of 600 g (51–75th percentile-for-gestational age) vaginally as a result of preterm labor, had multiple extreme prematurity-related problems, including anemia, hypotension, grade IV intraventricular hemorrhage, renal dysfunction, patent ductus arteriosis, pulmonary interstitial emphysema of the newborn, thrombocytopenia, neonatal hypocalcemia followed by hypercalcemia, hyperbilirubinemia of prematurity, hypo- and hyperglycemia and electrolyte abnormalities – initially hypernatremia accompanied by eucalemia, followed by hyponatremia and hyperkalemia during the last 24 h of life. Newborn screening 17-OH progesterone level on the second day of life was reported to be ‘within normal limits’, and a total T4 was reported as <3.0µg/dL, whereas the TSH was reported to be
2.3 μU/mL. The infant died on the 10th day of life of the complications of extreme prematurity and, in accordance with parental request, an autopsy was not done. The other two pregnancies concluded with spontaneous abortions. Because of the prior spontaneous abortions, and the extremely premature birth of the sibling, the mother of the proband received weekly injections of 17-hydroxyprogesterone caproate from 16 to 35 weeks of gestation. The last injection of 17-hydroxyprogesterone caproate (7) was received 1 week before the proband was born.

The proband infant had a prolapse of the upper vaginal region into the vaginal introitus with no visible secretions, partial labial-scrotal fusion, a 5 mm wide phallic structure with the urethral opening at the base, and no palpable gonads in the labial-scrotal folds (Fig. 1B). A chromosome analysis was ordered because of ambiguity of the genitalia (Fig. 1B). The infant was noted to have generalized hyperpigmentation, even when considering the Mexican ethnicity of the parents. The chromosome analysis revealed a 46, XY karyotype. On the sixth day of life, adrenocorticotropic hormone (ACTH) level was 3975 pg/mL (5–46 pg/mL), cortisol level was undetectable (<0.10 ng/mL), 17-hydroxyprogesterone was <10 ng/dL (29–35 weeks gestational age normal range 26–568 ng/dL), dihydroepiandrosterone (DHEA) was <0.050 ng/mL (premature normal range <40 ng/mL), androstenedione was <0.030 ng/mL (31–35 weeks gestational age normal range 0.80–4.46 ng/mL). The chromosome analysis, ACTH, plasma renin activity, steroids, with the exception of the cortisol, and their reference ranges are from ARUP LABORATORIES (Way, Salt Lake City, UT, USA). The serum glucose level was 58 mg/dL, sodium was 130 mmol/L, and the potassium was 5.7 mmol/L, and the infant was noted to be tachypneic and to have subcostal retractions. Stress doses of hydrocortisone sodium succinate were started.

When the tachypnea and retractions resolved, the serum sodium, potassium and glucose levels were stably within the normal ranges for age, the hydrocortisone sodium succinate was replaced with physiologic replacement doses of prednisone, as well as 9-alpha-fludrocortisone and sodium chloride supplementation. Since then, the infant’s linear growth and rate of weight gain have been normal, and the upper portion of the vagina is no longer protruding from the vaginal introitus. The adrenal gland was observed to be enlarged. The parents have opted to raise the child as a female unless the child opts for a male gender at an older age.

Investigation

Study participants: Informed consent was obtained from each participating family member as well as the parents of the patient with putative CAH.

Sequence analysis: Blood samples collected in ethylenediaminetetraacetic acid were used to prepare genomic DNA using a genomic DNA isolation kit (DNeasy Blood and Tissue Kit, Qiagen). Because the patient was considered to have lipid CAH, seven exons of StAR gene were analyzed following a standard procedure (3). Analysis of the CYP11A1 gene was also undertaken by first amplifying the genomic DNA using a variety of primer combinations (8), where we first amplified exons 1 and 2, which was similar to wild-type CYP11A1. We then amplified the rest of the exons using identical PCR conditions as previously described (8). PCR products were sequenced directly using commercial resources (MC Lab, South San Francisco, CA, USA).

Restriction fragment length polymorphism (RFLP) analysis: After the amplification of exon 5 using genomic DNA with primers SCCS5 and SCCSAS as previously described (8), the resultant 282bp product was incubated with BfaI at 37°C for 45 min. The bands were resolved in a non-denaturing 8% TBE acrylamide gel after electrophoresis at 60V for 2h and were visualized after staining with ethidium bromide.

Cell culture, transfection and activity analysis: COS-1 cells were grown in DMEM medium supplemented with 5% fetal bovine serum, 10% horse serum and 1× gentamycin at 37°C in a humidified incubator under 5% CO2. For CYP11A1 expression and activity, the mutant or wild-type CYP11A1 subcloned in pCMV vector and the purified plasmid cDNA was transfected with Lipofectamine as previously described (3). The cells were washed with serum-free media 12h after transfection and supplemented with media containing the appropriate antibiotics and 10% serum. In all cases, 100 ng/mL of trilostane (Steraloids, Los Angeles, CA, USA) was added as an inhibitor of 3βHSD2.

Metabolic conversion of cholesterol to pregnenolone was performed with reactions initiated with NADPH and 14C-cholesterol for pregnenolone synthesis. For complete conversion, we used five-fold excess of cold water-soluble cholesterol carrier to reach the saturation point. Trilostane was added in all the reactions to inhibit 3βHSD2 activity. Equal amounts of cold cholesterol-pregnenolone in ethanol were added as a carrier; the steroids were extracted with ether/acetone (9:1 v/v), and the solvent was evaporated under N2.
It was completed with the addition of five-fold excess of cold pregnenolone. The extract was brought up in 100µL of methanol, and radioactive counts were determined. The extracts were concentrated under a stream of nitrogen or air and then separated by TLC (Whatman, MA, USA) using a chloroform/ethyl acetate (3:1) mobile phase.

Mitochondrial translocation assay: A cell-free transcription/translation kit (Promega), which contains rabbit reticulocyte lysate (RRL) for protein expression after our modified procedure (6), was used to express wild-type and W286X CYP11A1-mutant proteins for in vitro experiments. Briefly, a mixture of 19 amino acids without cold methionine was incubated with 35S-methionine, (MP Biomedicals, Santa Ana, CA, USA) and SP6 polymerase at 26°C for 2h. Expression was verified using SDS-PAGE. For mitochondrial import analysis, mitochondria isolated from MA-10 cells were suspended in 20 mM HEPES, pH 7.5, following a previously described procedure (3, 4, 5, 6). After addition of the freshly prepared cell-free synthesized (CFS) protein of interest, mitochondria were incubated at 26°C. To inhibit the mitochondrial proton pump, 1.0 µg/mL valinomycin (Sigma) was added.

**Outcome and follow-up**

We have performed a pedigree analysis of the patient’s family, which shows that all the grandparents were from the same village and of the same ethnic origin (Fig. 1A). The grandparents lived a very long life with the paternal side living close to 100 years or more and the maternal side living more than 95 years. The parent’s cousins were all living, and no major genetic defect, physical disability or abnormality was known to the parents of the patient.

We first considered that the ambiguous female genitalia and adrenal insufficiency were possibly due to a mutation in the 7.5 kb StAR gene, which has seven exons with exon 5–7 comprising 40% of the protein. Although mutations have been identified in other exons, most StAR mutations causing lipoid CAH have been found in exons 5–7 of the StAR gene (3). Because we did not find any mutation in exons 4–7, we sequenced the complete gene but found no nucleotide changes in the StAR coding sequence even in the exon and intron boundary.

After revisiting the phenotypic data and finding that the mother had a problem retaining pregnancy, suggesting an inadequate supply of progesterone, we next analyzed all nine exons of the CYP11A1 gene using a combination of primers used previously (8). We identified a change in nucleotide 857, which changed from guanine to adenine, mutating tryptophan 286 to a stop codon (Fig. 2A). To confirm the presence of the mutation, we next amplified exon 5, including its intron–exon boundaries with the appropriate primers and performed RFLP analysis with BfaI because a TGG-to-TAG nucleotide change would destroy a BsrI restriction site at position 286 and simultaneously create a new BfaI restriction site (Fig. 2B and C). As shown in Fig. 3A, RFLP analysis using PCR-amplified product of exon 5 (282bp) from the patient, mother and father confirmed the presence of the mutation, resulting in two bands of 202 and 80bp in the patient. The wild-type product was not digested at all because of the absence of the BfaI restriction site. However, both of the parents showed a similar, partial
digestion with the intensities of the digested bands being 50% less as compared with the patient’s sample, showing that both parents were carriers. The RFLP analysis is presented schematically (Fig. 3A), including the position of the exon with its intronic boundaries (Fig. 3B).

After overexpression of the wild-type and mutant CYP11A1 in non-steroidogenic COS-1 cells that were also transfected with ferrodoxin, ferrodoxin reductase and P450scc as a single fusion protein, F2 (3), analysis of pregnenolone synthesis revealed that the wild-type vector synthesized 8-fold more pregnenolone compared with the mutant CYP11A1 (Fig. 3C and D), suggesting that W286X truncated CYP11A1 reduced progesterone synthesis but not to that level of complete inactivity.

To understand whether the inactivity of the mutant was due to the shorter protein sequence and/or whether it was incapable of interaction with mitochondrial translocases and, therefore, mitochondrial translocation, we compared the mitochondrial import of the W286X CYP11A1 mutant with wild-type CYP11A1 using radiolabeled CYP11A1 obtained via cell-free rabbit reticulocyte system (CFS) and mitochondria isolated from the steroidogenic mouse Leydig (MA-10) cells. As shown in Fig. 4A, the 61-kDa protein is imported into the mitochondria in two successive steps, resulting in 57 and 54 kDa proteins. The 61 and 54 kDa proteins were quickly proteolyzed by proteinase K (PK) in the presence of Triton X-100; however, the 54-kDa protein was resistant, suggesting that it was protected by the inner mitochondrial environment (Fig. 4A, last lane). Triton X-100 forms elliptical micelles on the membrane and makes an easy entry of the protease resulting in faster proteolysis. CYP11A1 was imported into the mitochondrial matrix as a 54 kDa protein, so a shorter incubation of PK and Triton X-100 together protected mostly the protein fragment imported into the matrix (Fig. 4A, Right hand Panel). Interestingly, the 31-kDa W286X mutant was also imported into the mitochondria and processed into a shorter 27-kDa band (Fig. 4B), suggesting that the CYP11A1 signal sequence is sufficient for mitochondrial transport. However, the regions required for interaction with the cofactors necessary for performing the enzymatic action may be lost in the truncated protein. To confirm the accuracy of the mitochondrial import and processing assay, we confirmed the quality of the mitochondria by importing cell-free synthesized CYP11A1 into the isolated mitochondria with the inhibition of the mitochondrial proton pump after incubation with 1.0 μg/mL valinomycin (Fig. 4C). CYP11A1 import was inhibited by valinomycin as indicated by the presence of only the 61 and 57 kDa proteins, thus validating our mitochondrial import and processing analysis.

Discussion

Progesterone synthesis by mitochondrial 3βHSD2 (2, 3, 4) is required for the maintenance of human pregnancies as it suppresses uterine contractility, and interruption in the
maintenance of progesterone level induces abortion. As Miller's group has considered, no mutation of the placental ferrodoxin, ferrodoxin reductase and 3βHSD2 was reported and none of the START domain proteins can compensate for the loss of CYP11A1 activity by cleaving cholesterol to pregnenolone. Thus, we analyzed whether a mutation in the CYP11A1 gene was the reason for the extremely high ACTH levels and PRA activity in the newborn and why the patient's mother experienced difficulty in retaining pregnancies. Our report demonstrates that the presence of homozygous disruptive CYP11A1 mutations may represent a preemptive indication for multiple spontaneous abortion. The mutation identified in this study was isolated from a Mexican newborn whose maternal and paternal families were from the same village and whose families had long and healthy lifespans.

To date, CYP11A1 gene mutations that induce CAH have been described in only 15 patients (Table 1), which is likely due to the fact that it is the sole enzyme maintaining adrenal activity required for the production of all steroids. Most CYP11A1 gene mutations have been heterozygous and some have appeared at the late onset of puberty. In contrast, we have identified a single nucleotide change at position 857 of the CYP11A1 gene, which results in a nonsense mutation (W286X) and, therefore, a truncated protein (from a 61 to a 31 kDa protein). The patient had very high ACTH levels and PRA activity, and the ACTH level remained higher than normal levels possibly because the mineralocorticoid receptor is not acting at the same level as the glucocorticoid receptor. As the ACTH level remained above the normal range, increased cortisol should then be maintained by the adrenal gland. However, inactivity of the metabolizing enzymes of the adrenal gland resulted in an imbalance in the electrolyte transport from the adrenal gland.

Many of the patients with previously identified CYP11A1 mutations had mothers with a history of miscarriages, and some experienced the very early stage of salt wasting crisis and high ACTH secretion (9). In some cases, the onset of adrenal failure was later, and many were
Table 1 Mutations in the side chain cleavage enzyme (CYP11A1) causing ambiguous genitalia, sex reversal, past miscarriages and spontaneous abortions.

<table>
<thead>
<tr>
<th>Mutations</th>
<th>Ref</th>
<th>Homozygous/ heterozygous (patient)</th>
<th>Onset of adrenal failure</th>
<th>Karyotype</th>
<th>Genitalia</th>
<th>Maternal miscarriage/ abortion</th>
<th>Gestation</th>
</tr>
</thead>
<tbody>
<tr>
<td>L222P</td>
<td>(3)</td>
<td>Homozygous</td>
<td>9 years</td>
<td>46, XY</td>
<td>Normal male</td>
<td>0</td>
<td>Full term</td>
</tr>
<tr>
<td>R360W/R405X</td>
<td>(3)</td>
<td>Heterozygous</td>
<td>15 days</td>
<td>46, XY</td>
<td>Micropenis</td>
<td>1</td>
<td>Full term</td>
</tr>
<tr>
<td>L141W/V415E</td>
<td>(3)</td>
<td>Heterozygous</td>
<td>1–9 days</td>
<td>46, XY</td>
<td>Normal female</td>
<td>0</td>
<td>Full term</td>
</tr>
<tr>
<td>c835delA</td>
<td>(3)</td>
<td>Homozygous</td>
<td>9 days</td>
<td>46, XY</td>
<td>Normal female</td>
<td>Low E₂ 2</td>
<td>31 weeks</td>
</tr>
<tr>
<td>Frameshift; c835delA stop/slice</td>
<td>(3)</td>
<td>Heterozygous</td>
<td>8 days</td>
<td>46, XY</td>
<td>Normal female</td>
<td>Low E₂ 1</td>
<td>Full term</td>
</tr>
<tr>
<td>R353W</td>
<td>(3)</td>
<td>Heterozygous</td>
<td>7–9 months</td>
<td>46, XY</td>
<td>Normal female</td>
<td>0</td>
<td>Full term</td>
</tr>
<tr>
<td>A189V</td>
<td>(3)</td>
<td>Heterozygous</td>
<td>7–9 months</td>
<td>46, XY</td>
<td>Normal female</td>
<td>0</td>
<td>Full term</td>
</tr>
<tr>
<td>A359V</td>
<td>(3)</td>
<td>Heterozygous</td>
<td>21 months</td>
<td>46, XY</td>
<td>Normal female</td>
<td>2</td>
<td>Full term</td>
</tr>
<tr>
<td>R232X</td>
<td>(10)</td>
<td>Homozygous</td>
<td>Neonate</td>
<td>46, XX</td>
<td>Normal female</td>
<td>2</td>
<td>Post-term</td>
</tr>
<tr>
<td>R232X</td>
<td>(10)</td>
<td>Homozygous</td>
<td>Neonate</td>
<td>46, XX</td>
<td>Normal female</td>
<td>0</td>
<td>Full term</td>
</tr>
<tr>
<td>R232X/F215S</td>
<td>(10)</td>
<td>Heterozygous</td>
<td>1.2 years</td>
<td>46, XY</td>
<td>Small penis</td>
<td>0</td>
<td>Twin at full term</td>
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<td>R232X/F215S</td>
<td>(10)</td>
<td>Heterozygous</td>
<td>4.75 years</td>
<td>46, XY</td>
<td>Normal male</td>
<td>0</td>
<td>Full term</td>
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<tr>
<td>R232X/F215S</td>
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<td>Heterozygous</td>
<td>1.5 years</td>
<td>46, XX</td>
<td>Normal female</td>
<td>0</td>
<td>Full term</td>
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<td>R120X</td>
<td>(10)</td>
<td>Homozygous</td>
<td>Neonate</td>
<td>46, XX</td>
<td>Normal female</td>
<td>1</td>
<td>Full term</td>
</tr>
<tr>
<td>A269V</td>
<td>(3)</td>
<td>Heterozygous</td>
<td>8 years</td>
<td>46, XY</td>
<td>Normal male</td>
<td>0</td>
<td>Full term</td>
</tr>
<tr>
<td>A269V</td>
<td>(3)</td>
<td>Heterozygous</td>
<td>12 months</td>
<td>46, XX</td>
<td>Normal female</td>
<td>0</td>
<td>Full term</td>
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<tr>
<td>R451W</td>
<td>(3)</td>
<td>Heterozygous</td>
<td>2 year 10 months</td>
<td>46, XY</td>
<td>Normal male</td>
<td>0</td>
<td>Full term</td>
</tr>
<tr>
<td>R451W</td>
<td>(3)</td>
<td>Heterozygous</td>
<td>1 year 3 months</td>
<td>46, XY</td>
<td>Normal male</td>
<td>0</td>
<td>Full term</td>
</tr>
<tr>
<td>Insertion of G and N between D271 and V272 W286X</td>
<td>(3)</td>
<td>Heterozygous</td>
<td>4 years</td>
<td>46, XY</td>
<td>Normal female</td>
<td>0</td>
<td>Full term</td>
</tr>
</tbody>
</table>

either heterozygotes or had undergone gene conversion, likely resulting in a minimum level of pregnenolone-to-progesterone conversion catalyzed by 3βHSD2 (4, 10).

Our recent finding shows that steroidogenic protein 3βHSD2 activity requires an association with mitochondrial translocases, including Tim50 or Tim23 of the inner mitochondrial translocase and Tom22 translocase facing the IMS side of the mitochondria, after entering to the mitochondria (10). In the absence of these mitochondrial translocases, 3βHSD2 and StAR activities were reduced. In the absence of the mitochondrial import receptor, Tom22, the alternate OMM-associated protein, Tom20, can facilitate protein import; however, it cannot translocate to the appropriate protein environment and thus becomes inactive. CYP11A1 is a 521 amino acid protein and its N-terminal 39 amino acids are essential for import into the mitochondria (3). In this study, the 286 amino acid CYP11A1 is a truncated protein, which is imported into the mitochondria where its 39 amino acid N-terminal signal sequence is cleaved in a fashion similar to the wild-type protein. Because of the absence of full-length sequence, the CYP11A1 passenger protein sequence could not interact with the electron donating partner of ferrodoxin – ferrodoxin reductase. Thus, the short CYP11A1 is imported into the mitochondria and processed in a similar fashion like wild type, which was not observed for another mitochondrial enzyme 3βHSD2, which ignored the N-terminal sequence when fused with its N-terminus (10). Thus, CYP11A1 activity requires the passenger protein sequence, which explains why carriers of the CYP11A1 mutation had retained activity.

Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

Funding
The work was initially supported by a grant from National Institutes of Health (HD057876), an intramural research grant from Anderson Cancer Institute and a Seed Grant from the Mercer University School of Medicine (all to H S B).

Patient consent
Informed consent has been obtained for this study.

Author contribution statement
J Kaur did the lab/scientific experiments and analyzed the data. H Bose analyzed and interpreted the data and wrote the manuscript. Patient was seen by A Rice, E O’Connor, A Piya, B Buckler.
CAH due to a novel CYP11A1 mutation

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Received in final form 7 September 2016
Accepted 13 September 2016