A surprising treatment response in a patient with rare isolated growth hormone deficiency, type IB

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Summary

Isolated Growth Hormone Deficiency (IGHD) is a rare cause of short stature, treated with the standard regimen of subcutaneous synthetic growth hormone (GH). Patients typically achieve a maximum height velocity in the first year of treatment, which then tapers shortly after treatment is stopped. We report a case of a 9-year-old male who presented with short stature (<3rd percentile for age and race). Basal hormone levels showed undetectable serum IGF1. Skeletal wrist age was consistent with chronologic age. Cranial MRI revealed no masses or lesions. Provocative arginine-GH stimulation testing demonstrated a peak GH level of 1.4 ng/mL. Confirmatory genetic testing revealed a rare autosomal recessive single-nucleotide polymorphism (SNP) with mutational frequency of 2%. GH supplementation was started and pursued for 2 years, producing dramatically increased height velocity. This velocity persisted linearly through adolescence, several years after treatment had been discontinued. Final adult height was >95th percentile for age and race. In conclusion, this is a case of primary hypopituitarism with differential diagnosis of IGHD vs Idiopathic Short Stature vs Constitutional Growth Delay. This case supports two objectives: Firstly, it highlights the importance of confirmatory genetic testing in patients with suspected, though diagnostically uncertain, IGHD. Secondly, it demonstrates a novel secondary growth pattern with implications for better understanding the tremendous variability of GH treatment response.

Learning points:

- GHD is a common cause of growth retardation, and IGHD is a specific subtype of GHD in which patients present solely with short stature.
- The standard treatment for IGHD is subcutaneous synthetic GH until mid-parental height is reached, with peak height velocity attained in the 1st year of treatment in the vast majority of patients.
- Genetic testing should be strongly considered in cases of diagnostic uncertainty prior to initiating treatment.
- Future investigations of GH treatment response that stratify by gene and specific mutation will help guide treatment decisions.
- Response to treatment in patients with IGHD is variable, with some patients demonstrating little to no response, while others are ‘super-responders.’
Background

GHD is a well-documented cause of growth retardation in both males and females, and affects 1/3800 children in the USA. It typically occurs with panhypopituitarism, presenting with low growth rate, hypoglycemia, microphallus, lethargy, and congenital blindness (1, 2). IGHD is a rare, specific subtype with deficiency only in GH; therefore, patients present exclusively with short stature. The majority of cases are idiopathic, though 3–30% of cases have shown traceable familial etiology. Mainstay treatment is subcutaneous synthetic GH injection until mid-parental height is reached. Because of the variability in clinical presentation and neurohormonal profile, clinical differentiation can be difficult. Further, the response to therapy ranges dramatically between patients. This case supports the use of confirmatory genetic testing in patients with suspected IGHD. Moreover, response to treatment is a key element to better understanding the pathogenesis of IGHD.

Case presentation

A 9-year-old male of Middle Eastern descent presented with a complaint of short stature relative to his peers. His mother reported that in a school with 87 males his age (as determined by school year), he was the shortest in stature. He otherwise had no history of developmental delay, reaching all developmental milestones at appropriate times (responding to name at 6 months, pulling-to-stand and waving ‘bye’ at 9 months, walking at 12 months). There was no history of birth injury (though he experienced a right inguinal hernia of his small intestine at 11 days of age that was surgically repaired; no complications of the surgery were reported) or significant head injury. Genealogically, his parents were not consanguineous. His mother's pregnancy was uneventful (birth weight: 3.62 kg, 71st percentile). Teacher reports described the patient’s behavior as ‘boisterous’ but without behavioral misconduct.

On examination, the patient’s height was 121.5 cm (3rd percentile for age and race). He showed no gross abnormalities on systemic examination. There were no dysmorphic features. He had no deficits in vision, motor coordination, or sensorineural hearing. Testicular volume (4.3 mL in each testes) and descent were normal. Head circumference was 51 cm. The ratio of upper:lower segment span (1.12) suggested that his short stature was proportionate for males his age. Examination of secondary sexual characteristics revealed Tanner Stage II. Penis length was age-appropriate (4.5 cm).

Investigation

X-ray bone age studies of the left wrist were consistent with chronologic age and race. Initial screening with serum Insulin Growth Factor 1 (IGF1) was abnormally low (24.1 ng/mL). Basal Serum GH was also low (0.02 ng/mL). IGF Binding Protein 3 (IGF-BP3) was within normal limits (4.6 ng/L). Testosterone (6.4 ng/dL) and sex hormone-binding globulin (SHBG; 89 nmol/L) were also within normal reference range. Adrenal function tests, thyroid function tests, liver function tests, and hemoglobin A1C were all normal (Table 1). 2-Plane cranial MRI showed no abnormalities. Mid-parental height was 150.25 cm. Growth velocity from age 8 to 9 was determined as 1.3 cm/year. Provocative arginine-GH stimulation testing was undertaken during a 12-h fasting period. A standard in-house sex steroid priming protocol was followed prior to provocative testing as the patient’s exam suggested pre-pubescent developmental stage. GH stimulation testing revealed a peak GH level of 1.4 ng/mL (lower limit of normal: 10 ng/mL). Confirmatory molecular testing was sent for identification of possible mutations or copy-number repeats in genes GH1 and GHRH, and transcription factors LHX3, LHX4 and PROP1, FOU1F1.

Table 1 Baseline endocrine data.

<table>
<thead>
<tr>
<th>Test</th>
<th>Unit</th>
<th>Values</th>
<th>Reference Range</th>
</tr>
</thead>
<tbody>
<tr>
<td>HbA1C</td>
<td>%</td>
<td>4.6</td>
<td>4.3–5.8</td>
</tr>
<tr>
<td>TSH</td>
<td>μU/mL</td>
<td>2.2</td>
<td>0.34–3.5</td>
</tr>
<tr>
<td>fT4</td>
<td>ng/dL</td>
<td>1.2</td>
<td>0.9–1.8</td>
</tr>
<tr>
<td>ACTH</td>
<td>pg/mL</td>
<td>2.4</td>
<td>2.0–4.0</td>
</tr>
<tr>
<td>Cortisol</td>
<td>mg/dL</td>
<td>8.1</td>
<td>4.4–52</td>
</tr>
<tr>
<td>Basal GH</td>
<td>ng/mL</td>
<td>0.02</td>
<td>0–20</td>
</tr>
<tr>
<td>Peak GH</td>
<td>ng/mL</td>
<td>1.4</td>
<td>&gt;5</td>
</tr>
<tr>
<td>IGF1</td>
<td>ng/mL</td>
<td>24</td>
<td>68–216</td>
</tr>
<tr>
<td>IGF-BP3</td>
<td>mg/L</td>
<td>4.6</td>
<td>1.8–7.1</td>
</tr>
<tr>
<td>LH</td>
<td>IU/mL</td>
<td>5.1</td>
<td>1.6–9.5</td>
</tr>
<tr>
<td>PRL</td>
<td>ng/mL</td>
<td>1.3</td>
<td>1.2–15</td>
</tr>
<tr>
<td>Test</td>
<td>ng/dL</td>
<td>6.4</td>
<td>&lt;7–20</td>
</tr>
<tr>
<td>SHBG</td>
<td>nmol/L</td>
<td>89</td>
<td>49–179</td>
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<tr>
<td>NSB</td>
<td>%</td>
<td>0.4</td>
<td>&lt;8</td>
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<tr>
<td>FIRI</td>
<td>μU/mL</td>
<td>4.8</td>
<td>1.7–10.4</td>
</tr>
<tr>
<td>TIRI</td>
<td>μU/mL</td>
<td>5.6</td>
<td>5.0–15</td>
</tr>
</tbody>
</table>

ACTH, adrenocorticotropic hormone; FIRI, free immunoreactive insulin; FSH, follicle stimulating hormone; fT3, free triiodothyronine; fT4, free thyroxine; GH, growth hormone; HbA1c, hemoglobin A1c; IGF1, insulin-like growth factor-I; LH, luteinizing hormone; NSB, non-specific binding; PRL, prolactin; peak GH, peak stimulated growth hormone; SHBG, sex hormone-binding globulin; test, testosterone; TIRI, total immunoreactive insulin; TSH, thyroid stimulating hormone.
and HESX1. Mutational analysis revealed a 5′IVS-4 single nucleotide (Guanine-to-Cytosine) nonsense mutation in the 17q23.3 locus. A literature review indicated that this mutation has been attributed exclusively to IGHD, type IB (3). Accordingly, the patient was diagnosed with this specific subtype. Additional genetic testing was performed in the parents and living grandparents, which identified each parent and 2 grandparents as heterozygotic carriers. The paternal grandfather was unavailable for testing. The maternal grandmother was deceased.

**Treatment**

After obtaining parental and patient consent, once-daily subcutaneous administration of synthetic GH was started at age 9 years and 322 days, and was discontinued 412 days later. During that time, dosage was titrated to weight (0.18 mg/kg/week) at every three-month visit. Serum IGF1 was measured at these intervals to verify that dosing remained within a clinically therapeutic range. At no point during the treatment window did IGF1 exceed the age-adjusted upper limit of normal. Repeat IGF1 screening at ages 14 (102 ng/mL) and 15 (132 ng/mL) were also within appropriate range.

**Outcome and follow-up**

Body height increased to 152.7 cm (62nd percentile) by the end of the treatment period (Fig. 1). Body weight increased from 26.7 kg (33rd percentile) to 41.6 kg (50th percentile). Height velocity during the 1st year of treatment was 8.1 cm/year. Treatment was stopped shortly after age 12, when the mid-parental height was reached. The patient’s height on the last day of treatment was 150.3 cm (51st percentile). Following cessation of treatment, rapid linear growth continued into adulthood (Fig. 1). Final adult height was 186.8 cm (96th percentile). Final weight at this point in time was 72.4 kg (66th percentile). Sexual maturation continued at age-appropriate milestones (Tanner Stage 3: 12.6 years, Tanner Stage 4: 14.5 years, Tanner Stage 5: 15.7 years). Height velocities during the first and second year following treatment were 10.4 and 11.1 cm/year, respectively. Height velocities for the 5 consecutive years following cessation of treatment were 11.8, 10.7, and 9.2, 7.3, and 4.5 cm/year, respectively (Fig. 2).

**Discussion**

This case highlights a rare cause of primary hypopituitarism with a broad differential diagnosis. Idiopathic Short Stature was considered as a possible cause, but was unlikely as the patient demonstrated undetectable levels of serum IGF-1 on repeat testing prior to beginning therapy. Direct pituitary damage (e.g. traumatic brain injury, Pituitary Apoplexy) was considered as possible etiologies in retrospective analyses. However, there was no history of trauma, birth complications, or ingestion of toxic substances, and MRI revealed no corresponding lesion. We considered the possibility of unrecognized maternal alcohol consumption, but it is unlikely since the patient did not have any of the pathognomonic facial features of Fetal Alcohol Syndrome (low nasal bridge, epicanthal folds, and micrognathia). Moreover, alcohol consumption has only been identified in the literature as a cause of panhypopituitarism rather than IGHD (4). Suspection of
neurological damage secondary to maternal ingestion of other toxins was low as the patient demonstrated no other physical abnormalities or cognitive deficits on evaluation, both of which are typical sequela (5). Nutritional deficit was not suspected given serum albumin and pre-albumin levels were within normal reference ranges and proportionate weight change prior to treatment.

Lastly, given the singular complaint of short stature, we considered Constitutional Growth Delay. It is well-known that up to 70% of children with isolated GHD will not have persistent GHD into adolescence or adulthood (6). Accordingly, we remained wary of initiating GH treatment without clear indication. Patients with Constitutional Growth Delay tend to resume normal height velocities by ages 2–3 and a delayed skeletal age (as estimated by wrist X-ray), commonly 2–4 years behind chronological age (7, 8). Our patient demonstrated low height velocity through age 9, and his skeletal age was consistent with chronological age. However, these trends are by no means definitive. Moreover, both IGH and Constitutional Growth Delay may show poor response to provocative testing (9), calling into question the inter-trial reliability of testing (10). Given the clinical equipoise, we elected for genetic testing. The 5′IVS-4 single nucleotide (Guanine-to-Cytosine) nonsense SNP has a mutational frequency of ~2% and presents with mild-to-severe short stature without dysmorphies or neuroendocrine derangements (3).

IGHD, type IB is characterized by autosomal recessive inheritance, a severely diminished endogenous GH pattern, and a very low but detectable peak GH stimulatory test (11). This endocrine profile matched that of our patient. Moreover, genetic testing of the parents identified a phylogenetically traceable, heterogenous mutation in each parent, indicative of autosomal inheritance. In this particular case, genetic testing allowed for definitive diagnosis of IGH, type IB. While some clinicians may argue in favor of close monitoring of height velocity for a self-correcting growth delay, mathematical modeling has identified age of treatment onset as one of the strongest predictors of treatment response (12). Delaying treatment would have possibly diminished the patient’s therapeutic outcome. Recent work by Khadilkar et al. showed that in children matched for age and baseline height, treatment response to GH in those with homozygous GH1 deletions was poor relative to those with a GHRHR mutation or without any known mutation (13). In cases of previously-identified ‘poor responders,’ genetic testing may save families from ultimately fruitless and expensive therapies. Often, screening of the parents is sufficient, though in our patient’s case, we elected to pursue additional family members due to the rarity of this particular mutation.

Our own patient represents the other end of the spectrum of the treatment response: a ‘super-responder.’ Though type IB is typically a severe form of GHD, treatment response with lower levels of GH show reasonable response, while avoiding development of anti-GH antibodies (14).

However, treatment response can vary significantly and is not well-characterized (2, 14). These potential limitations were discussed with the family, who elected to proceed regardless. While reasonable to expect some efficacy, the phenomenon observed in our patient was fairly surprising. We considered that this sustained growth rate may simply be a pubertal surge. However, even in healthy males, growth continues asymptotically as of age 15 at a rate roughly half of that observed in our patient (15). Our patient’s observed height velocities far exceeded the standardized height velocities, which were first characterized by Backer et al. (16), and validated by others (17, 18). Moreover, to our knowledge, there is no other reported case of IGHD of any subtype where height velocity after treatment exceeded that of the 1st year of treatment.

Based on these surprising observations, we conclude that the range of treatment response in patients with IGHD may vary far more than previously understood. With an increasing number of source mutations being identified, future investigations of treatment efficacy would likely benefit from greater stratification by genetic mutation.

Declaration of interest
All authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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Patient consent
Informed patient consent was obtained for publication in this journal. Our patient was a legal adult at the time of manuscript preparation, and was able to sign consent for himself.
Patient's perspective

Our patient declined to submit a patient perspective, but he did communicate to us that reviewing our work and his case inspired his own desire to pursue a career in translational research.

Author contribution statement

J A was responsible for drafting the manuscript and data synthesis. K B was responsible for drafting the manuscript and literature review. E W was responsible for creating the code used to generate figures and literature review. T A was the supervising physician responsible for clinical accuracy and reviewing the manuscript.

References


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