Identification of a novel large \textit{CASR} deletion in a patient with familial hypocalciuric hypercalcemia

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Summary

Familial hypocalciuric hypercalcemia type I is an autosomal dominant disorder caused by heterozygous loss-of-function mutations in the \textit{CASR} gene and is characterized by moderately elevated serum calcium concentrations, low urinary calcium excretion and inappropriately normal or mildly elevated parathyroid hormone (PTH) concentrations. We performed a clinical and genetic characterization of one patient suspected of familial hypocalciuric hypercalcemia type I. Patient presented persistent hypercalcemia with normal PTH and 25-hydroxyvitamin D levels. The \textit{CASR} was screened for mutations by PCR followed by direct Sanger sequencing and, in order to detect large deletions or duplications, multiplex ligation-dependent probe amplification (MLPA) was used. One large deletion of 973 nucleotides in heterozygous state (c.1733-255_2450del) was detected. This is the first large deletion detected by the MLPA technique in the \textit{CASR} gene.

Learning points:

- Molecular studies are important to confirm the differential diagnosis of FHH from primary hyperparathyroidism.
- Large deletions or duplications in the \textit{CASR} gene can be detected by the MLPA technique.
- Understanding the functional impact of the mutations is critical for leading pharmacological research and could facilitate the therapy of patients.

Background

Familial hypocalciuric hypercalcemia type I (FHH1, OMIM \#145980) is an autosomal dominant disorder transmitted with a high degree of penetrance and characterized by moderately elevated serum calcium concentrations, low urinary calcium excretion and inappropriately normal or mildly elevated parathyroid hormone (PTH) concentrations. This disorder is caused by heterozygous loss-of-function mutations in the \textit{CASR} gene (OMIM +601199) encoding the human calcium-sensing receptor (CaSR).

Multiple disorders have been associated with defects in the CaSR as it plays a central role in the regulation of extracellular calcium homeostasis (1, 2). FHH1, neonatal hyperparathyroidism (NHPT, OMIM \#239200) and hypercalciuric hypercalcemia (3) are diseases caused by heterozygous loss-of-function mutations in the \textit{CASR} gene, whereas activating mutations can result in isolated autosomal dominant hypocalcemia (OMIM \#601198) or combined with Bartter-like syndrome (OMIM \#601198).
The aim of this study was to perform a clinical and genetic characterization of one patient suspected of familial hypocalciuric hypercalcemia. Patients with mutations of the CASR gene may not present such a classic picture of hypercalcemia with hypocalciuria or hypocalcemia with hypercalcuria. Molecular studies are important for confirming the diagnosis and distinguish it from other entities. One large deletion of 973 nucleotides in heterozygous state (c.1733-255_2450del) was detected in our patient. Importantly, as far as we know, this is the first large deletion detected by multiplex ligation-dependent probe amplification (MLPA) technique in the CASR gene.

**Case presentation**

The study was approved by the Ethics Committee for Clinical Research of Euskadi (CEIC-E). The patient is a 56-year-old female of Philippine origin who was referred for evaluation of persistent hypercalcemia in several routine blood laboratory analyses. A history of hypercalcemia was reported in her family as well (her sister showed hypercalcemia and hyperparathyroidism, she does not have a genetic diagnosis yet). Unfortunately, we do not have any information about the parents. Laboratory results showed persistent hypercalcemia (serum calcium of 11.6 mg/dL, reference range 8.5–10.5 and total serum calcium corrected for albumin of 11.7 mg/dL, reference range 8.6–10.2) and serum phosphate at the lower limit of the normal range (2.8 mg/dL, reference range 2.6–4.8), whereas intact PTH (40 pg/mL, reference range 10–65) and 25-hydroxyvitamin D levels (34.4 ng/mL, reference range 8.6–54.8) were within the normal range. The patient exhibited hypocalciuria. Specifically, her urinary calcium was very low (urinary calcium <5 mg/dL, reference range 0–250; urinary creatinine 35 mg/dL, reference range 90–300) despite their high serum calcium levels, and she had a 24-h urine calcium:creatinine clearance ratio (CCCR) <0.01. Therefore, the patient was suspected of FHH and the CASR gene genetics analysis was requested. Four years previously, she had developed hyperthyroidism due to Graves–Basedow disease, and she had been treated with antithyroid medications and radiiodine therapy. Furthermore, the patient presented other personal history as hypercholesterolemia, prediabetes and femoral and lumbar osteoporosis (markers of bone remodeling: serum alkaline phosphatase 60 U/L, reference range 46–116 U/L; serum osteocalcin 3.02 ng/mL, reference range 0–22; β-isomerized C-telopeptides 0.12 µg/L, reference range 0–0.84).

**Investigation**

In order to detect large deletions or duplications in the CASR region, a commercially available MLPA kit, SALSA MLPA probemix P177-B2 (MRC Holland, Amsterdam, The Netherlands), was used. This MLPA kit contains 14 MLPA probes specific for the CASR gene with amplification products between 130 and 301 nucleotides. In addition, it contains nine reference probes expected to have a normal copy number that we used for the intrasample normalization, and nine internal quality control fragments generating an amplification product smaller than 120 nucleotides.

The exons and flanking intronic sequences of the CASR gene (Ensembl identifiers: gene ENSG00000036828; transcript, ENST00000639785.1) were screened for mutations by PCR followed by direct sequencing. We used primers to amplify exons 2–7 together with their splice sites. Furthermore, we designed primers to amplify a large fragment where the large deletion, found by the MLPA technique, is included (Forward: 5’-ACAGGTAAGGAAACCCCTCTT-3’; Reverse: 5’GCCTCCACCACTGATGACAA-3’). Purified amplified products were directly sequenced in both directions with fluorescent dideoxynucleotides (BigDye Terminator v3.1 Cycle Sequencing Kit, Life Technologies). DNA mutation was named according to the Human Genome Variation Society guidelines (www.hgvs.org).

With respect to genotype, one large deletion of 973 nucleotides in heterozygous state (marked in bold in the CASR sequence showed in *Table 1*) was detected by the MLPA technique (Fig. 1). The amplification of exon 7 using specific primers that jointed to the deleted region confirmed the heterozygous state in the patient. Subsequently, to delimit the deletion, we amplified one fragment of 2926 nucleotides by long-PCR and delimited the deletion to 973 nucleotides by sequencing with internal primers. The deletion expands from nucleotide in c.1733-255 position within intron 6 to nucleotide c.2518 within exon 7 (c.1733-255_2450del). This deletion is supposed to generate a protein shorter than WT, as the splice site of exon 7 is affected. Therefore, the mutant CaSR protein probably loses the transmembrane domain and the intracellular carboxyl-terminal tail (amino acids 578 to 1078).

As far as we know, this is the first large deletion detected by MLPA technique in the CASR gene.
Table 1: Gross deletion marked in bold within the CASR gene sequence (Ensembl identifiers: gene ENSG00000036828; transcript, ENST00000639785.1).

<table>
<thead>
<tr>
<th>Exon</th>
<th>Sequence</th>
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<tbody>
<tr>
<td>6</td>
<td>GTGCCCTTCTCCAACTGCAGCCGAGACTGCCTGGCAGGGACCAGGAAAGGGATCATTGAGGGGGAGCCCACCTGCTGCTTTGAGTGTGTGGAGTGTCCTGATGGGGAGTATAGTGATGAGACAGGAGTCAGGGAACCTT</td>
</tr>
<tr>
<td>Intron 6-7</td>
<td>gtagaagggaacccctcttgggcactgtgcagggcttggtccacttcggaggcctggggtcagtgaagcccatggaagggctgggctgagatggtgctttgcacagtttctcatttaacaaaggtatatctgagcatcacaaaatgttgggcctca</td>
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Gross deletion marked in bold within the gene sequence (Ensembl identifiers: gene ENSG00000036828; transcript, ENST00000639785.1).

Exon 7

<table>
<thead>
<tr>
<th>Exon</th>
<th>Sequence</th>
</tr>
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<tbody>
<tr>
<td>7</td>
<td>ATGCCAGTGCCTGTAACAAGTGCCCAGATGACTTCTGGTCCAATGAGAACCACCTCCTGCATTGCCAAGGAGATCGAGTTTCTGTCGTGGACGGAGCCCTTTGGGATCGCACTCACCCTCTTTG</td>
</tr>
</tbody>
</table>

Discussion

In this study, we have described the first large deletion detected in the CASR gene found in the homozygous state. The patient is affected by a severe form of hereditary hypocalcemia. The deletion involves the splice site of exon 7 in a gene where loss of function is a known mechanism of disease; consequently, this large deletion has very strong evidence of pathogenicity; therefore, it is classified as class 4 (pathogenic) by the ACMG guidelines.

We hypothesized that our patient only presents CASR WT mutations; a true WT receptor signal. We confirmed that in addition, we found that the patient possesses a high degree of similarity to the WT receptor signal.

As the splice site of exon 7 is included in the deleted region, it is supposed that the mutant protein lacks both the transmembrane and the intracellular domain. Thus, the allele presenting the deletion produces a non-functional and thus, does not interfere with the WT receptor signal.
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Presenting the deletion. The second deletion consisted in a large deletion of chromosome 3p13.31-22.1 that included 101 known genes, including the CASR gene. The individual presented multiple disorders, including hypercalcemia and hypocalciuria (9).

Identification of large deletions by MLPA is remarkably rare in the CASR gene. Thus, we only identified one copy number variation in 56 patients (2%) with calcium metabolism disorders using the MLPA technique (data not published). Moreover, a previous study did not identify any copy number variation in a large cohort of 257 patient samples using MLPA (9, 10). In our laboratory, firstly, the exons and flanking intronic sequences of the CASR gene are screened for mutations by PCR followed by direct sequencing, and secondly, when the results are negative, the MLPA is performed. This technique is a useful tool to check copy number variations (CNVs). In our experience, the next-generation sequencing shows great sensitivity but little specificity in CNVs detection. Therefore, we propose to first look for mutations by sequencing, should mutation not be detected, MLPA or QMPSF (quantitative multiplex PCR of short fluorescent) analyses must be carried out in order to complete the molecular study.

In conclusion, molecular studies are important to confirm the differential diagnosis of FHH from primary hyperparathyroidism (thus preventing surgery) and other diseases, because of common biochemical features. Understanding the functional impact of the mutations at the cell membrane is critical for leading pharmacological research and could facilitate the therapy of patients. Moreover, performing genetic analysis is essential to detect carriers, because some individuals with mutations remain normocalcemic. The aim of this study was to describe the first gross deletion found using the MLPA technique. Punctual mutations are the most common variants described in the CASR gene (according to the Human Gene Mutation Database, 348 mutations). We demonstrated that MLPA is a useful technique for the detection of large deletions or duplications in the CASR gene. Therefore, we recommended the use of this technique to complete the molecular study.

Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of this case report.

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Patient consent
The patient provided written, informed consent to participate in this study. The information has been sufficiently anonymized, making it impossible to identify the patient with any certainty.
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Author contribution statement

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References

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